

Understanding the Role of the Endoplasmic Reticulum Stress Sensor IRE1 α in Aging

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Abstract: Aging is a progressive biological process that leads to a decline in multiple organ functions and finally to disease. A key feature of aging is the breakdown of proteostasis, particularly affecting the endoplasmic reticulum (ER). As an adaptation to ER stress, the unfolded protein response (UPR) is activated to maintain ER homeostasis. Among the three arms of the UPR, IRE1 α is the most ancient pathway involved in maintaining proteostasis. As such, multiple studies demonstrate IRE1 α 's significance in ensuring healthy aging. The process of aging can be delayed through various strategies, including genetic modifications, dietary adjustments, and pharmacological interventions, which are correlated with improved ER proteostasis, activation of the UPR and IRE1 α . This article reviews the latest progress in understanding the role of the UPR and IRE1 α signaling in the aging process and its interconnection with other aging biomarkers, including cellular senescence.

Keywords: UPR, ER Proteostasis, Healthy Aging

1. Introduction

With the aging population increasing around the world, deciphering the molecular pathways and processes that lead to aging is becoming more critical. The inability to give an adequate response to cellular stress is toxic in aging organisms, resulting in accumulated damage to cells and loss of proteostasis. Also, age-dependent unfolded protein accumulation during aging drives aging-related diseases. The ability of organisms to respond to ER stress, also known as the UPR (unfolded protein response), decreases with aging, which is harmful in the face of accumulated unfolded proteins, indicating the importance of the UPR in resolving ER stress (Endoplasmic Reticulum Stress) in aging [1, 2].

The Endoplasmic reticulum (ER) is a membrane-bound organelle that is crucial for protein synthesis, folding, and trafficking within cells. Upon disruption of cellular homeostasis (e.g., hypoxia, aging, physical exercise, infections, glucose or calcium imbalance, etc.), the demand for protein synthesis increases. If newly synthesized peptides cannot be folded properly in a timely manner, the accumulation of unfolded proteins will induce ER stress (Endoplasmic Reticulum Stress), triggering the activation of the UPR (unfolded protein response) [3-5]. There are three ER-resident transmembrane sensors of the UPR: protein kinase R (PKR)-like endoplasmic reticulum kinase (PERK), inositol-requiring enzyme 1 (IRE1), and activating transcription factor 6 (ATF6) [6]. Among them, IRE1 is the most evolutionarily conserved UPR sensor [7].

IRE1 α is a transmembrane protein comprising a luminal domain, a linker region, and a cytoplasmic domain [8]. During the resting state, BiP/glucose-regulated protein 78 (GRP78), an ER-resident chaperone, interacts with the luminal domain of IRE1 α to restrain its activity [5]. Loss of ER homeostasis causes BiP to dissociate from IRE1 α , leading to the activation of IRE1 α . Besides, the binding of unfolded proteins to IRE1 α can also induce its activation [9]. Upon activation, IRE1 α undergoes autophosphorylation via its kinase activity and oligomerization to induce its RNase activity, which is located in its cytoplasmic tail. RNase activation of IRE1 α initiates unconventional splicing of XBP1 mRNA by cleaving out 26 nucleotides, followed by ligation by a tRNA ligase, RTCB, to generate the spliced form of XBP1 (XBP1s). XBP1s then enters the nucleus to induce the expression of UPR-related genes, including those encoding protein chaperones, ERAD (ER-associated degradation) components,

and genes involved in ER biogenesis, to resolve ER stress [10, 11]. Besides XBP1, the cleavage targets of the RNase activity of IRE1 α can include microRNAs [12] and mRNAs, a process termed regulated IRE1 α -dependent decay (RIDD) [2, 13, 14]. In addition to its RNase-dependent functions, IRE1 α can also participate in the JNK and IKK β -NF- κ B inflammatory signaling pathways via its interaction with TNF-receptor-associated factor 2 (TRAF2) [15, 16].

2. The role of IRE1 α in aging

2.1 IRE1 α in aging in simple model organisms

The components of the UPR, especially IRE1 α , are vital for longevity in simple animal models, including fruit flies, *C. elegans* and yeast. Yeast cells have a shorter lifespan due to mutations within the IRE1 α pathway, which is the only UPR pathway in yeast [17]. IRE1 α plays an important role in longevity partly through its splice target, the spliced form of XBP1 (XBP1s). Overexpression of XBP1s in the small intestinal cells of fruit flies can extend their lifespan, while overexpression of XBP1s in the fat body of fruit flies can promote longevity by regulating their metabolic state [18]. Besides, the unspliced form of XBP1 (XBP1u) also plays a role in lifespan extension [18], although the detailed mechanism remains to be elucidated. The protection conferred by XBP1s to fruit flies from Alzheimer's disease decreases with aging, consistent with the decreasing intensity of the UPR observed with aging [19].

C. elegans has been widely used to elucidate mechanisms involved in longevity. The significance of the IRE1/XBP1 pathway in neurons for healthy aging has been established [11, 20-24]. Both overexpression and loss-of-function (lof) mutations of XBP1s can affect the lifespan of *C. elegans* significantly. Specifically, XBP1s exerts its lifespan extension function by inducing the UPR in the intestine via secreting small clear vesicles [20]. Overexpression of XBP1s in the intestine of *C. elegans* demonstrated that the induced UPR in the gut regulates genes involved in lipophagy [22], proteostasis [23] and lipid metabolism [24]. Overexpression of XBP1s in different types of neurons exerts different effects on changes in gene expression patterns in peripheral tissues. Whereas the UPR in dopaminergic neurons modulates lipid depletion distally, the UPR in serotonergic neurons plays a role in promoting adaptive homeostasis throughout the body [25]. Overexpression of XBP1s in glutamatergic neurons enhances protein homeostasis, whereas overexpression of XBP1s in octopaminergic neurons upregulates the antipathogen immune response [26]. Meanwhile, activated UPR in GABAergic neurons has less effect on lipid metabolism homeostasis [26]. It remains unclear why expression of XBP1s in different neuronal types has different effects.

As important components of the nervous system, glial cells, in addition to providing protection, maintain metabolic homeostasis of neurons. Overexpression of XBP1s in the four astrocyte-like glial cells of *C. elegans* promotes longevity through modulating proteostasis throughout the body [21]. The expression of TFEB, a transcription factor that regulates lysosome biosynthesis, is upregulated when XBP1s is overexpressed in glial cells to regulate lipid metabolism and intestinal autophagy [27]. Induced UPR in neuronal and glial cells can regulate gut and whole-body proteostasis in a cell-nonautonomous manner. When the mitochondrial UPR is activated, astrocyte-like glial cells can also send signals to neurons through small clear vesicles and to peripheral tissues through dense-core vesicles to promote longevity [28].

The Nervous system of *C. elegans* can sense ER stress to send signals to peripheral tissues to modulate global adaptation and thereby regulate lifespan. In addition to ER stress, the heat shock response has a similar function. Activation of IRE1/XBP1s by volatile pathogen-associated compounds in neuronal cells can promote lifespan [29]. In summary, activation of IRE1/XBP1s to induce cellular adaptation to ER stress can benefit health and extend lifespan.

2.2 IRE1 α in aging in mammalian aging

During healthy aging, ER proteostasis is impaired in human tissues. Increased UPR in the eye lens indicates increased ER stress [30], and this increased ER stress is related to functional defects in aging islet β cells [31]. Meanwhile, ER stress in human muscle attenuates with aging, a process that can be improved by physical exercise [32]. Higher levels of ER stress and aging markers were detected in lung fibroblasts derived from older humans than from younger humans [33]. Thus, the intensity of the UPR decreases with aging, impairing its ability to maintain ER proteostasis, which may result in accumulation of misfolded/unfolded proteins and damage to ER proteostasis.

Mouse models provide a powerful tool for manipulating gene expression to study the UPR in mammalian aging. Overexpression of XBP1s in neurons can delay or even prevent cognitive decline in healthy aging [34]. 70% of the proteomic changes observed in the brain during normal aging can be reversed by overexpression of XBP1s in the hippocampus of mice, accompanied by improved synaptic function [34]. Genes upregulated by overexpression of XBP1s, which are downregulated in the hippocampus with healthy aging in mice, are also downregulated in the hippocampus of aged humans [34].

Conditional knockout of IRE1 α in the retina results in decreased outer nuclear layer thickness as animals age [35], indicating the significance of the IRE1/XBP1s pathway in ocular aging.

The IRE1/XBP1s pathway has also been linked to ocular aging. In a model of retinitis pigmentosa, which is related to chronic ER stress, mice were more susceptible to retinal degeneration due to defective UPR [35]. A model with deletion of XBP1s in the retina exhibits dysfunction of photoreceptors [36]. Thus, whether IRE1/XBP1s can regulate lifespan in mammals remains to be determined.

2.3 IRE1 α in cell senescence

Although loss of ER proteostasis is a sign of the aging process, UPR signaling can promote other aspects of aging. Cellular senescence is characterized by stable growth arrest, a proinflammatory secretome and other changes [37]. Human senescent cells show enhanced ER stress sensing, but insufficient UPR-related transcriptional induction [38]. The activation of the IRE1/XBP1s pathway is important for cell senescence caused by DNA damage [39].

The UPR activation and enhanced autophagy in senescent cells are involved in the upregulation of SASP (senescence-associated secretory phenotype) secretion [40-42]. DNA damage agent-induced senescence increased ER stress when autophagy was inhibited with quercetin, leading to senolysis [43]. Conditional knockout of ATF6 or IRE1 in β cells of non-obese diabetic (NOD) mice prior to insulinitis induces a senescence phenotype in β cells [44]. The IRE1/XBP1s pathway is involved in regulating the number of senescent cells in the brain during aging. Whereas deletion of IRE1 α accelerates the accumulation of senescent cells during aging, overexpression of XBP1s reduces the number of senescent cells [34]. IRE1 α regulates the response to DNA damage through RIDD, which can lead to a senescent phenotype; conversely, DNA damage can also trigger the oligomerization of IRE1 α to catalyze RIDD [45].

Verteporfin treatment, which inhibits the YAP-TEAD pathway, reduces ER biogenesis and induces ER stress [46]. Owing to high protein secretion demand imposed by the SASP, the activated UPR is required to maintain ER proteostasis to sustain cell survival. Consequently, VPF treatment can selectively reduce the viability of senescent cells [46]. Thus, the high demand for protein synthesis and secretion in senescent cells underlies the significance of the UPR in regulating senescence.

2.4 IRE1 α as a mediator in aging-modifier approaches

Multiple interventions to extend lifespan act through the IRE1/XBP1s pathway. Under dietary restriction, the IRE1/XBP1s pathway is activated in a metabolic adaptation that promotes enterocyte homeostasis and longevity in *Drosophila* [47]. The IRE1/XBP1s pathway can also enhance the lifespan of insulin/IGF-1 signaling mutant *C. elegans* [48]. Interaction between XBP1s and the FOXO transcription factor can promote longevity in a fruit fly model [18]. IRE1/XBP1s is required for the longevity effects of dietary restriction, resulting in increased ER-associated degradation (ERAD) gene expression and degradation of ER proteins [49].

Exposure of *C. elegans* to Tm (tunicamycin, an ER stress activator) in early development can enhance proteostasis, prevent age-related UPR decline, and extend adult lifespan [49], a phenomenon called ER hormesis. ER preconditioning using genetic manipulation to alter ER homeostasis can also promote longevity [50]. Glucose and vitamin D restriction can also promote longevity of *C. elegans* via a UPR-dependent pathway [51, 52].

Endurance exercise can promote healthy aging at least partly through inducing metabolic stress and by improving ER proteostasis [53]. Endurance exercise can prevent aging-induced chronic ER stress in muscle [54]. Endurance exercise induces less upregulation of the UPR gene expression program in older humans [32], possibly due to impaired UPR activation. Resistance training and swimming can restrict the expression of ER stress-related proteins in the heart during aging [55, 56]. Resistance training can also induce the UPR program in peripheral blood mononuclear cells from older humans [57].

Metformin, an AMPK activator, can prevent aging-related hearing loss and reduce the UPR activation [58]. ER stress declines and cognitive function improves in mice after Metformin intake [59]. Although a correlation between the UPR and Metformin has been established, direct evidence is still needed.

3. Discussion

An organism's robustness and the preservation of its health rely on homeostatic mechanisms that respond to molecular changes, orchestrating reactions across organs. It is anticipated that the disruption of proteostasis during aging influences cells, since maintaining the cell-type-specific proteomes is the cornerstone of health maintenance. The molecular mechanism underlying the declined activity of the proteostasis network during aging remains an unresolved question, potentially stemming from a lack of evolutionary pressure to preserve proteome integrity after reproduction. The aging process may encompass increased impairment of ER proteostasis---manifesting as alterations in the secretory pathway, malfunction of the ER folding apparatus, protein aggregation within the ER, among others---or a deficiency in monitoring and repairing damage, such as the oxidative inactivation of UPR stress sensors, which could, in turn, exacerbate the breakdown of proteostasis.

We hypothesize that the ability to detect misfolded proteins becomes compromised with age, potentially leading to the decline of tissue integrity, based on published studies. The brain-gut axis is a dynamic and reciprocal communication system that functionally links different organs. While there is substantial evidence from invertebrate studies indicating a relationship between neuronal UPR and protein homeostasis in the gut, it remains unclear if such a link exists in mammals. This uncertainty poses a significant question for future research. Notably, research has demonstrated that XBP1s in the hypothalamus can induce a cell-nonautonomous activation of the UPR in the liver, influencing energy metabolism [60]. Nonetheless, the role of this pathway in the aging process has yet to be elucidated.

The exposome plays a pivotal role in shaping the aging process in humans. Our sensory perceptions act as the critical link, bridging our body with the external environment. The link between the perception of food, the hypothalamus, and hepatic XBP1s signaling might represent evidence of a sensory-proteostasis nexus in mammals [61]. As previously discussed, in *C. elegans*, the perception of olfactory cues exerts life-extending influences by modulating the UPR. Similarly, in these organisms, responses to cold stress have been associated with IRE1 signaling [62], a pathway that influences both longevity and health [63]. We conjecture that the nervous system functions as a central hub, collecting diverse information from the exposome and capable of discerning even the slightest deviations in cellular homeostasis. It then sends signals throughout the organism to foster adaptation and promote survival.

Despite the availability of sophisticated genetic tools that allow the manipulation of various UPR signaling pathways in specific tissues [11, 64], evidence connecting UPR signaling to the aging process in mammals remains insufficient, with the majority of research focusing on the nervous system. Enhancing the UPR artificially could potentially delay or even prevent the decline in tissue function associated with aging. We emphasize the potential widespread advantages of boosting UPR activity in the brain---for instance, through gene therapy---not only to enhance brain function but also to regulate proteostasis throughout the entire organism. In recent years, a surge in research has underscored the pivotal role of the UPR in promoting healthy aging, while also revealing numerous challenges. Given that biological age can diverge from chronological age at the level of individual organs and cell types, it remains to be determined whether interventions aimed at the UPR can alter biological age, alongside other key indicators of aging such as cellular senescence and inflammation.

The decline in ER proteostasis could serve as a prognostic indicator for aging patterns and as a biomarker to evaluate the effectiveness of potential therapeutic interventions. It has been suggested that the evaluation of aging clocks might incorporate the analysis of tissue-specific protein aggregation [65], based on the observation that aging leads to the aggregation of proteins. Proteomic analysis of plasma has revealed that markers of proteostasis undergo changes early in life, providing predictive insights into the likelihood of developing dementia [66], and that age-related changes specific to tissues may be identifiable in blood samples [67]. These insights open the door to testing the effectiveness of drugs targeting the UPR for aging-related interventions, as well as to monitoring biomarkers of aging for personalized medicine through ex vivo cultures. Similar to the approach taken with senolytics, we suggest that interventions aimed at improving ER proteostasis could mitigate a wide range of diseases, potentially due to the UPR's fundamental role in the biology of aging, which is a primary risk factor for the onset of numerous chronic diseases.

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References

- [1] Hetz, C. and A. Dillin, *Central role of the ER proteostasis network in healthy aging*. *Trends Cell Biol*, 2024.
- [2] Han, D., et al., *IRE1alpha kinase activation modes control alternate endoribonuclease outputs to determine divergent cell fates*. *Cell*, 2009. 138(3): p. 562-75.
- [3] Hetz, C. and L.H. Glimcher, *Protein homeostasis networks in physiology and disease*. *Curr Opin Cell Biol*, 2011. 23(2): p. 123-5.
- [4] Schröder, M. and R.J. Kaufman, *The mammalian unfolded protein response*. *Annu Rev Biochem*, 2005. 74: p. 739-89.
- [5] Walter, P. and D. Ron, *The unfolded protein response: from stress pathway to homeostatic regulation*. *Science*, 2011. 334(6059): p. 1081-6.
- [6] Acosta-Alvear, D., et al., *Homeostasis control in health and disease by the unfolded protein response*. *Nat Rev Mol Cell Biol*, 2024.
- [7] Junjappa, R.P., et al., *IRE1a Implications in Endoplasmic Reticulum Stress-Mediated Development and Pathogenesis of Autoimmune Diseases*. *Front Immunol*, 2018. 9: p. 1289.
- [8] Huang, S., Y. Xing, and Y. Liu, *Emerging roles for the ER stress sensor IRE1a in metabolic regulation and disease*. *J Biol Chem*, 2019. 294(49): p. 18726-18741.
- [9] Gardner, B.M. and P. Walter, *Unfolded proteins are Ire1-activating ligands that directly induce the unfolded protein response*. *Science*, 2011. 333(6051): p. 1891-4.
- [10] Lu, Y., F.X. Liang, and X. Wang, *A synthetic biology approach identifies the mammalian UPR RNA ligase RtcB*. *Mol Cell*, 2014. 55(5): p. 758-70.
- [11] Hetz, C., K. Zhang, and R.J. Kaufman, *Mechanisms, regulation and functions of the unfolded protein response*. *Nat Rev Mol Cell Biol*, 2020. 21(8): p. 421-438.
- [12] Chitnis, N., D. Pytel, and J.A. Diehl, *UPR-inducible miRNAs contribute to stressful situations*. *Trends Biochem Sci*, 2013. 38(9): p. 447-52.
- [13] Hollien, J., et al., *Regulated Ire1-dependent decay of messenger RNAs in mammalian cells*. *J Cell Biol*, 2009. 186(3): p. 323-31.
- [14] Hollien, J. and J.S. Weissman, *Decay of endoplasmic reticulum-localized mRNAs during the unfolded protein response*. *Science*, 2006. 313(5783): p. 104-7.
- [15] Urano, F., et al., *Coupling of stress in the ER to activation of JNK protein kinases by transmembrane protein kinase IRE1*. *Science*, 2000. 287(5453): p. 664-6.
- [16] Hu, P., et al., *Autocrine tumor necrosis factor alpha links endoplasmic reticulum stress to the membrane death receptor pathway through IRE1alpha-mediated NF-kappaB activation and down-regulation of TRAF2 expression*. *Mol Cell Biol*, 2006. 26(8): p. 3071-84.
- [17] Chadwick, S.R., et al., *A functional unfolded protein response is required for chronological aging in *Saccharomyces cerevisiae**. *Curr Genet*, 2020. 66(1): p. 263-277.
- [18] Li, M., et al., *Xbp1 targets canonical UPR(ER) and non-canonical pathways in separate tissues to promote longevity*. *iScience*, 2024. 27(6): p. 109962.
- [19] Marcora, M.S., et al., *Amyloid-β42 clearance and neuroprotection mediated by X-box binding protein 1 signaling decline with aging in the *Drosophila* brain*. *Neurobiol Aging*, 2017. 60: p. 57-70.
- [20] Taylor, R.C. and A. Dillin, *XBP-1 is a cell-nonautonomous regulator of stress resistance and longevity*. *Cell*, 2013. 153(7): p. 1435-47.
- [21] Frakes, A.E., et al., *Four glial cells regulate ER stress resistance and longevity via neuropeptide signaling in *C. elegans**. *Science*, 2020. 367(6476): p. 436-440.
- [22] Daniele, J.R., et al., *UPR(ER) promotes lipophagy independent of chaperones to extend life span*. *Sci Adv*, 2020. 6(1): p. eaaz1441.
- [23] Imanikia, S., et al., *Neuronal XBP-1 Activates Intestinal Lysosomes to Improve Proteostasis in *C. elegans**. *Curr Biol*, 2019. 29(14): p. 2322-2338.e7.
- [24] Imanikia, S., et al., *XBP-1 Remodels Lipid Metabolism to Extend Longevity*. *Cell Rep*, 2019. 28(3): p. 581-589.e4.
- [25] Higuchi-Sanabria, R., et al., *Divergent Nodes of Non-autonomous UPR(ER) Signaling through Serotonergic and Dopaminergic Neurons*. *Cell Rep*, 2020. 33(10): p. 108489.
- [26] Coakley, A.J., et al., *Distinct mechanisms of non-autonomous UPR(ER) mediated by GABAergic, glutamatergic, and octopaminergic neurons*. *bioRxiv*, 2024.

- [27] Metcalf, M.G., et al., *Cell non-autonomous control of autophagy and metabolism by glial cells*. *iScience*, 2024. 27(4): p. 109354.
- [28] Bar-Ziv, R., et al., *Glial-derived mitochondrial signals affect neuronal proteostasis and aging*. *Sci Adv*, 2023. 9(41): p. eadi1411.
- [29] De-Souza, E.A., M.A. Thompson, and R.C. Taylor, *Olfactory chemosensation extends lifespan through TGF- β signaling and UPR activation*. *Nat Aging*, 2023. 3(8): p. 938-947.
- [30] Tang, H.Z. and L.M. Yang, *Activation of the unfolded protein response in aged human lenses*. *Mol Med Rep*, 2015. 12(1): p. 389-93.
- [31] Shrestha, S., et al., *Aging compromises human islet beta cell function and identity by decreasing transcription factor activity and inducing ER stress*. *Sci Adv*, 2022. 8(40): p. eabo3932.
- [32] Hart, C.R., et al., *Attenuated activation of the unfolded protein response following exercise in skeletal muscle of older adults*. *Aging (Albany NY)*, 2019. 11(18): p. 7587-7604.
- [33] Koloko Ngassie, M.L., et al., *Endoplasmic reticulum stress-induced senescence in human lung fibroblasts*. *Am J Physiol Lung Cell Mol Physiol*, 2024. 327(1): p. L126-L139.
- [34] Cabral-Miranda, F., et al., *Unfolded protein response IRE1/XBP1 signaling is required for healthy mammalian brain aging*. *Embo j*, 2022. 41(22): p. e111952.
- [35] Massoudi, D., et al., *Deletion of the Unfolded Protein Response Transducer IRE1 α Is Detrimental to Aging Photoreceptors and to ER Stress-Mediated Retinal Degeneration*. *Invest Ophthalmol Vis Sci*, 2023. 64(4): p. 30.
- [36] McLaughlin, T., et al., *Loss of XBP1 Leads to Early-Onset Retinal Neurodegeneration in a Mouse Model of Type I Diabetes*. *J Clin Med*, 2019. 8(6).
- [37] Campisi, J., *Aging, cellular senescence, and cancer*. *Annu Rev Physiol*, 2013. 75: p. 685-705.
- [38] Sabath, N., et al., *Cellular proteostasis decline in human senescence*. *Proc Natl Acad Sci U S A*, 2020. 117(50): p. 31902-31913.
- [39] Blazanin, N., et al., *ER stress and distinct outputs of the IRE1 α RNase control proliferation and senescence in response to oncogenic Ras*. *Proc Natl Acad Sci U S A*, 2017. 114(37): p. 9900-9905.
- [40] Ei, Z.Z., et al., *GRP78/BiP determines senescence evasion cell fate after cisplatin-based chemotherapy*. *Sci Rep*, 2021. 11(1): p. 22448.
- [41] Ketkar, M., et al., *Inhibition of PERK-mediated unfolded protein response acts as a switch for reversal of residual senescence and as senolytic therapy in glioblastoma*. *Neuro Oncol*, 2024. 26(11): p. 2027-2043.
- [42] Dörr, J.R., et al., *Synthetic lethal metabolic targeting of cellular senescence in cancer therapy*. *Nature*, 2013. 501(7467): p. 421-5.
- [43] Bientinesi, E., et al., *Quercetin induces senolysis of doxorubicin-induced senescent fibroblasts by reducing autophagy, preventing their pro-tumour effect on osteosarcoma cells*. *Mech Ageing Dev*, 2024. 220: p. 111957.
- [44] Lee, H., et al., *Stress-induced β cell early senescence confers protection against type 1 diabetes*. *Cell Metab*, 2023. 35(12): p. 2200-2215.e9.
- [45] Dufey, E., et al., *Genotoxic stress triggers the activation of IRE1 α -dependent RNA decay to modulate the DNA damage response*. *Nat Commun*, 2020. 11(1): p. 2401.
- [46] Anerillas, C., et al., *The YAP-TEAD complex promotes senescent cell survival by lowering endoplasmic reticulum stress*. *Nat Aging*, 2023. 3(10): p. 1237-1250.
- [47] Luis, N.M., et al., *Intestinal IRE1 Is Required for Increased Triglyceride Metabolism and Longer Lifespan under Dietary Restriction*. *Cell Rep*, 2016. 17(5): p. 1207-1216.
- [48] Henis-Korenblit, S., et al., *Insulin/IGF-1 signaling mutants reprogram ER stress response regulators to promote longevity*. *Proc Natl Acad Sci U S A*, 2010. 107(21): p. 9730-5.
- [49] Matai, L., et al., *Dietary restriction improves proteostasis and increases life span through endoplasmic reticulum hormesis*. *Proc Natl Acad Sci U S A*, 2019. 116(35): p. 17383-17392.
- [50] Chinchankar, M.N., et al., *A novel endoplasmic reticulum adaptation is critical for the long-lived *Caenorhabditis elegans* rpn-10 proteasomal mutant*. *Biochim Biophys Acta Gene Regul Mech*, 2023. 1866(3): p. 194957.
- [51] Beaudoin-Chabot, C., et al., *The unfolded protein response reverses the effects of glucose on lifespan in chemically-sterilized *C. elegans**. *Nat Commun*, 2022. 13(1): p. 5889.
- [52] Mark, K.A., et al., *Vitamin D Promotes Protein Homeostasis and Longevity via the Stress Response Pathway Genes *skn-1*, *ire-1*, and *xbp-1**. *Cell Rep*, 2016. 17(5): p. 1227-1237.
- [53] Estébanez, B., et al., *Endoplasmic Reticulum Unfolded Protein Response, Aging and Exercise: An Update*. *Front Physiol*, 2018. 9: p. 1744.
- [54] Belaya, I., et al., *Long-Term Exercise Protects against Cellular Stresses in Aged Mice*. *Oxid Med Cell Longev*, 2018. 2018: p. 2894247.
- [55] Tang, J., et al., *Resistance training up-regulates *Smyd1* expression and inhibits oxidative stress and*

endoplasmic reticulum stress in the heart of middle-aged mice. *Free Radic Biol Med*, 2024. 210: p. 304-317.

[56] Chang, P., et al., Swimming exercise inhibits myocardial ER stress in the hearts of aged mice by enhancing cGMP-PKG signaling. *Mol Med Rep*, 2020. 21(2): p. 549-556.

[57] Estébanez, B., et al., Effects of a resistance-training programme on endoplasmic reticulum unfolded protein response and mitochondrial functions in PBMCs from elderly subjects. *Eur J Sport Sci*, 2019. 19(7): p. 931-940.

[58] Cai, H., et al., Metformin attenuates the D-galactose-induced aging process via the UPR through the AMPK/ERK1/2 signaling pathways. *Int J Mol Med*, 2020. 45(3): p. 715-730.

[59] Xu, X., et al., Metformin activates chaperone-mediated autophagy and improves disease pathologies in an Alzheimer disease mouse model. *Protein Cell*, 2021. 12(10): p. 769-787.

[60] Williams, K.W., et al., Xbp1s in Pomc neurons connects ER stress with energy balance and glucose homeostasis. *Cell Metab*, 2014. 20(3): p. 471-82.

[61] Brandt, C., et al., Food Perception Primes Hepatic ER Homeostasis via Melanocortin-Dependent Control of mTOR Activation. *Cell*, 2018. 175(5): p. 1321-1335.e20.

[62] Dudkevich, R., et al., Neuronal IRE-1 coordinates an organism-wide cold stress response by regulating fat metabolism. *Cell Rep*, 2022. 41(9): p. 111739.

[63] Lee, H.J., et al., Cold temperature extends longevity and prevents disease-related protein aggregation through PA28 γ -induced proteasomes. *Nat Aging*, 2023. 3(5): p. 546-566.

[64] Marciniak, S.J., J.E. Chambers, and D. Ron, Pharmacological targeting of endoplasmic reticulum stress in disease. *Nat Rev Drug Discov*, 2022. 21(2): p. 115-140.

[65] Dormann, D. and E.A. Lemke, Adding intrinsically disordered proteins to biological ageing clocks. *Nat Cell Biol*, 2024. 26(6): p. 851-858.

[66] Walker, K.A., et al., Proteomics analysis of plasma from middle-aged adults identifies protein markers of dementia risk in later life. *Sci Transl Med*, 2023. 15(705): p. eadf5681.

[67] Oh, H.S., et al., Organ aging signatures in the plasma proteome track health and disease. *Nature*, 2023. 624(7990): p. 164-172.